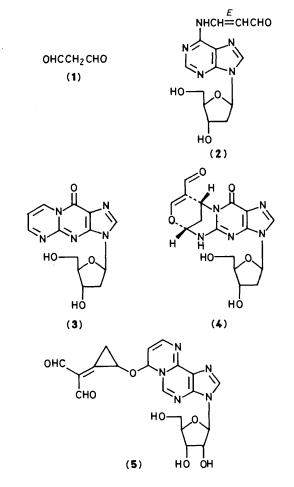
Dimer and Trimer of Malonaldehyde

Bernard T. Golding, ** Naina Patel, * and William P. Watson *

^a Department of Chemistry, Bedson Building, The University, Newcastle upon Tyne NE1 7RU ^b Shell Research Limited, Sittingbourne Research Centre, Sittingbourne, Kent, ME9 8AG

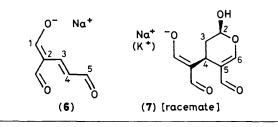
Self-condensation of malonaldehyde in water at pH 4.2 gives (E)-2-formylpent-3-ene-1,5-dial (**6**) and 4-(1,3-dioxopropan-2-yl)-3,4-dihydro-2H-pyran (**7**)

Malonaldehyde (propanedial) (1) can be formed *in vivo* by the oxidation of unsaturated lipids,¹ as a by-product of the biosynthesis of prostanoids,² and by attack of radicals on deoxyribose.³ It is mutagenic⁴ and is implicated in the ageing process.⁵ Consequently, there is considerable interest in the reactions of malonaldehyde with DNA and proteins. Recently, adducts of malonaldehyde with nucleosides⁶⁻⁸ and DNA⁹ have been described. These were 1:1 adducts⁶⁻⁸ [*e.g.* compounds (2) and (3)], 2:1 adducts⁷ [*e.g.* (4)], and 3:1 adducts⁸ [*e.g.* (5)].



In connection with our studies of cyclic nucleic acid adducts¹⁰ we have investigated the oligomerisation chemistry of malonaldehyde in aqueous solution. It has long been known that malonaldehyde oligomerises in water, but despite many studies no conclusive structural evidence for the oligomers has been presented.^{11–13} However, condensation products of malonaldehyde are known to react with DNA⁴ and proteins.¹⁴ To help assess the relevance of the oligomers to the toxicology of malonaldehyde, we have attempted to obtain these compounds pure and to characterise them. We have found that malonaldehyde undergoes a relatively rapid dimerisation and trimerisation at 20 °C in water at pH 4.2. We report herein the structures of the compounds formed and some preliminary information pertaining to their chemistry.

A 1M solution (pH 4.2) of malonaldehyde 15 in water was monitored by ¹H n.m.r. spectroscopy and this showed the formation of (E)-2-formylpent-3-ene-1,5-dial (6) and 4-(1,3dioxopropan-2-yl-5-formyl-2-hydroxy-3,4-dihydro-2H-pyran (7) (ratio 1:4) at 20 °C. These compounds had appeared within 15 min and all the malonaldehyde had been consumed after 3 days. Only after 25 days were additional products present in appreciable concentrations. From preparative scale reactions compounds (6) and (7) were isolated as metal salts by fractional crystallisation and chromatography.[†]

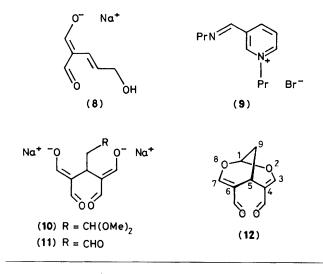


† Reactions initially contained either the potassium or sodium salt of malonaldehyde (0.3M in water). The pH was adjusted to 4.2 (N.B. pK_a of malonaldehyde = 4.46) by addition of dilute hydrochloric acid. After disappearance of the malonaldehyde (19 h at 37 °C) the solution was concentrated and sodium chloride was precipitated by addition of methanol. The filtrate was concentrated and the residue was redissolved in methanol. This solution when cooled to 0 °C and diluted with dichloromethane gave a precipitate of compound (7). This was further purified by recrystallisation from aqueous acetone to give a white solid, m.p. 161-162 °C (potassium salt). The filtrate from the initial precipitation of (7) was concentrated and the residue was chromatographed on silica (elution of the orange band with 15% methanol in dichloromethane) to give compound (6). This was recrystallised from methanol-dichloromethane to give an orange solid (decomposed without melting on heating). Both compounds (6) and (7) contained one sodium ion (or potassium ion, when the potassium salt of malonaldehyde was used as starting material) per molecule. For (6): δ_H(D₂O) 7.00 (1 H, dd, J 8.7 and 15.4 Hz, 4-H), 7.54 (1 H, d, J 15.4 Hz, 3-H), 8.95 (2 H, br s, 1-H and 2-CHO) and 9.25 (1 H, d, J 8.7 Hz). The ¹H n.m.r. spectrum of (6) in acidic D₂O gradually simplifies because of exchange of 4-H; δ_C(CD₃OD) 117.3 (C-3), 122.8 (C-4), 149.7 (C-2), 191.9 (C-1 and 2-CHO), and 198.8 (C-5); λ_{max} (H₂O) 275 (ϵ 5 000), 350 (6 500) and 476 nm (300); λ_{\max} (acidic H₂O) 260 (5 000) and 295 nm (4 300); ν_{\max} (KBr) 1 655 cm⁻¹; m/z (f.a.b.) 149 (corresponds to protonated monosodium salt). For (7): $\delta_{\rm H}$ 1.94 (2 H, dd, J 4 and 6 Hz, 2 × 3-H), 3.70 (1 H, t, J 6 Hz, 4-H), 5.50 (1 H, t, J 4 Hz, 2-H) and 8.20 (4 H, br s, 6-H, 5-CHO, and OCH=CCHO); λ_{max} (H₂O) 250 (ϵ 20 100) and 270 nm (12 200); λ_{max} (acidic water) 251 nm (31 900); v_{max} (KBr) 1 615 and 1 648 cm⁻¹; m/z (f.a.b.) 275 [corresponds to protonated dipotassium salt (11)].

Structure (6) follows from spectroscopic data * and the finding that addition of sodium borohydride to a solution of (6) in D_2O showed immediate formation of an alcohol assigned structure (8): δ_H 4.26 (2 H, d, J 6 Hz, 2 × 5-H), 6.44 (1 H, d, J 16 Hz, 3-H), 6.65 (1 H, dt, J 6 and 16 Hz, 4-H), 8.64 (2 H, br s, 1-H and 2-CHO).

Structure (7) follows from spectroscopic data,* including comparison with data for compound (10)[†] [prepared by the neutralisation with NaOH of a partial acidic hydrolysis of 1,1,3,3-tetramethoxypropane (*i.e.* condensation of 1 molecule of 3,3-dimethoxypropanal with 2 molecules of malonaldehyde, cf. ref. 16)]. The ¹H n.m.r. spectrum of compound (7) is pH dependent. Raising the pD of a solution of (7) in D_2O to 12 showed its conversion into the dianion (11), in which the aldehyde function can be trapped by borohydride reduction. Equilibration of (7) with (11) explains why the ^{1}H n.m.r. spectrum* of compound (7) shows apparent equivalence of 6-H, 5-CHO, and the protons in the OCH=CCHO group. Ring-opening of (7) to (11), can be followed by one of two energetically identical modes of cyclisation. The cis stereochemistry assigned to (7) is consistent with its acid-catalysed cyclisation to 2,8-dioxabicyclo[3.3.1]nona-3,6-diene-4,6-dicarbaldehyde (12). Thus, adjustment to pH 2 of a solution of (7) or (10), and extraction with dichloromethane gave the acetal (12), m.p. 148 °C, $\delta_{\rm H}$ (CDCl₃) 1.49 (2 H, dd, J 2 and 2 Hz, 2 × 9-H), 3.68 (1 H, dt, 2 and 5 Hz, 5-H), 5.77 (1 H, dt, 2 and 5 Hz, 1-H), 6.94 (2 H, s, 3-H and 7-H), and 8.88 (2 H, s, 2 × CHO).†

The 2:1 and 3:1 adducts described 7,8 from reactions of malonaldehyde and nucleosides can either be explained as



* See footnote on p. 268.

⁺ Satisfactory spectroscopic and analytical data were obtained for this compound.

products of a stepwise condensation of a nucleoside with 2 (or 3) molecules of malonaldehyde, or could arise from the condensation of (6) or possibly (7) with a nucleoside. The rate of self-condensation of malonaldehyde is competitive with its rate of condensation with nucleosides.^{8.17} We are therefore investigating reactions of (6) and (7) with nucleosides and oligonucleotides, although no experimentation is required to conclude that the structures and mechanisms proposed⁸ for the 3:1 adducts of malonaldehyde with nucleosides are unlikely! The potential ability of (6) to crosslink oligonucleotides and/or proteins was explored in a model experiment. Thus, reaction of 1.35M (6) with propylamine hydrobromide (2 mol equiv.) in D₂O at pD 10.2 gave the propylamine imine of N-propyl-3formylpyridinium bromide (9) within 30 min at 20 °C. This substance[†] was separately prepared by treating propylamine with N-propyl-3-formylpyridinium bromide[†] (obtained by treating pyridine-3-carbaldehyde with 1-bromopropane).

Acknowledgements

We thank S.E.R.C. for an award to N. P., Dr. S. J. Hill for n.m.r. spectra and Shell for use of laboratory facilities.

References

- 1 H. Esterbauer in 'Aldehydic Products of Lipid Oxidation,' ed. D. C. H. McBrien and T. F. Slater, Academic Press, New York, 1982, pp. 101–128.
- 2 M. Hamberg and B. Samuelsson, J. Biol. Chem., 1967, 242, 5344.
- 3 R. M. Burger, A. R. Berkowitz, J. Peisach, and S. B. Horwitz, *J. Biol. Chem.*, 1980, **255**, 11832.
- 4 A. K. Basu, L. J. Marnett, and L. J. Romano, *Mutation Res.*, 1984, 129, 39.
- 5 M. Rothstein, Chem. Eng. News, 1986, Aug. 11, p. 26.
- 6 R. C. Moschel and N. J. Leonard, J. Org. Chem., 1976, 41, 294; H. Seto, T. Takesue, and T. Ikemura, Bull. Chem. Soc., Jpn., 1985, 58, 3431.
- 7 L. J. Marnett, A. K. Basu, S. M. O'Hara, P. E. Weller, A. F. M. M. Rahman, and J. P. Oliver, *J. Am. Chem. Soc.*, 1986, **108**, 1348; A. K. Basu, S. M. O'Hara, P. Valladier, K. Stone, O. Mols, and L. J. Marnett, *Chem. Res. Toxicol.*, 1988, **1**, 53.
- 8 V. Nair, G. A. Turner, and R. J. Offerman, J. Am. Chem. Soc., 1984, 106, 3370.
- 9 H. Seto, T. Seto, T. Okuda, T. Takesue, and T. Ikemura, *Chem. Pharm. Bull.*, 1986, **34**, 5079.
- 10 B. T. Golding, P. K. Slaich, and W. P. Watson, J. Chem. Soc., Chem. Commun., 1986, 515.
- 11 J. M. C. Gutteridge, Anal. Biochem., 1975, 69, 518.
- 12 H. Buttkus, J. Agric. Food Chem., 1975, 23, 823.
- 13 H. Heusinger, Carbohydrate Res., 1986, 154, 37.
- 14 B. C. Shin, J. W. Huggins, and K. L. Carraway, Lipids, 1972, 7, 229.
- 15 R. Huttel, Chem. Ber., 1941, 74, 1825.
- 16 F. Wille and W. Schwab, Monatsh. Chem., 1977, 108, 929.
- 17 B. T. Golding, N. Patel, and W. P. Watson, unpublished results.

Received 12th July 1988

(Accepted 22 November 1988); Paper 8/02714E